

ESSENTIAL OIL MODIFICATION OF GLASS IONOMER CEMENT: ANTIBACTERIAL ACTIVITY AND COMPRESSIVE STRENGTH

Maha Ebrahim Elkorashy* 

ABSTRACT

Objective: The current study was conducted to investigate the effect of essential oil (EO) modification of glass ionomer cement (GIC) on antibacterial activity and compressive strength.

Methods: A total of 100 specimens were prepared; 50 specimens for the antibacterial activity test (3 mm x 2 mm) and 50 specimens for the compressive strength test (4 mm x 6 mm). Specimens were divided into five groups (n=10) according to the type of the essential oil used in modification of GIC; Group 1: unmodified glass ionomer cement (control group); Group 2: 2.5% bergamot modified GIC; Group 3: 2.5% peppermint modified GIC Group 4: 2.5% lemongrass modified GIC and Group 5: 2.5 % fennel modified GIC. Antibacterial activity was assessed using agar diffusion test. Compressive strength test was performed using universal testing machine. Data were tabulated and statistically analyzed.

Results: Bergamot modified GIC recorded the highest mean inhibition zone, followed by fennel, then lemongrass and peppermint. The lowest mean value was revealed by the control group. Regarding compressive strength results, control group showed the highest mean value, followed by lemongrass, then peppermint and bergamot. The lowest mean value was recorded by fennel modified GIC.

Conclusion: Essential oil modification of GIC was effective in enhancing the antibacterial activity although it adversely affects compressive strength in comparison to the control group. Lemongrass and peppermint essential oil modification of GIC were effective in improving antibacterial property while preserving an acceptable compressive strength according to the ISO standards.

KEY WORDS: Essential oil, Bergamot, Peppermint, Lemongrass, Fennel

* Associate Professor, Operative Dentistry Department Faculty of Dentistry Fayoum University Egypt

INTRODUCTION

Dental caries is a multi-factorial disease that occurs as a result of ecological imbalance between protective mechanisms and oral microbial biofilms. It causes destruction of dental hard tissues by bacterial acids due to fermentation of dietary carbohydrates¹. *Streptococcus mutans* plays the main role in development of dental caries as it is predominantly isolated from initial and advanced carious lesions².

Microbial biofilms are controlled using different mechanical and chemical methods for plaque control. Mechanical plaque control could be held either by mastication as natural mechanism or by using tooth brushes, dental floss and fissure sealants³. One of the most frequent approaches used for chemical plaque control is the use of antibacterial agents⁴.

Restorative materials with antibacterial properties are highly recommended. Fluoride release capacity of glass ionomer cements supports its anticariogenic potential⁵. However, the bacterial spectrum that could be inhibited due to fluoride release is still limited⁶. Addition of various synthetic constituents to glass ionomer cements was proposed in an attempt to enhance its antibacterial activity as chloroxylenol⁶, chlorhexidine gluconate⁷, doxycycline hyclate⁸, tetracycline⁹, ciprofloxacin, metronidazole and minocycline¹⁰.

Increasing microbial resistance to antibiotics has promoted developing natural antibiotics of plant origin as a next-generation for synthetic antibiotics⁹. Essential oils (EOs) are naturally derived components found in flowers, fruits, herbs, seeds, leaves and roots of different plants¹¹. They consist mainly of mixture of volatile compounds which are classified in two main groups; terpenes and terpenoids as well as aromatic and aliphatic constituents, in addition to other components in lower concentrations¹². Essential oils have antibacterial potential via destroying bacterial cell

walls and disrupting their enzymatic activity¹³.

Bergamot is a hybrid fruit between bitter orange and lemon¹⁴. Its essential oil is extracted from the peel of the *Citrus bergamia* Risso et Poiteau plant. It is reported to have valuable biological activities including antifungal, anti-inflammatory¹⁵ as well as antibacterial activity against *mutans streptococci*¹⁴. Peppermint is a medicinal plant that also known as *Mentha piperita* from the family of Lamiaceae. Menthol is the main component of peppermint. It possesses an antibacterial activity against *streptococcus mutans*¹⁶. Lemongrass (*Cymbogon citratus*) is a plant that belongs to the Poaceae family. It possesses antimicrobial, anti-rheumatic, analgesic, antispasmodic and antipyretic properties¹⁷. It was previously reported to have an antibacterial effect against *streptococcus mutans*⁹. Fennel or *Foeniculum vulgare* is a medicinal plant from the family of Apiaceae. It has anti-inflammatory, antioxidant¹⁶ and antibacterial potential¹⁸.

Although, several researchers reported an antibacterial activity of the previously mentioned essential oils against *streptococcus mutans*^{9,13,14,16}, no researches assessed the effect of modification of glass ionomer cement using bergamot, peppermint, lemongrass and fennel on antibacterial activity.

Glass-ionomer cement (GIC) is a water-based material that sets depending on acid-base reaction. This setting reaction is a slowly progressing process associated with a gradual increase in material mechanical properties¹⁹. Dental restorations should have acceptable mechanical properties as they are subjected to various stresses inside the oral cavity during functional loading as compressive, tensile and shear stresses²⁰. Compressive strength test is advocated by the ISO standards for evaluating the mechanical properties of dental restorations because most of the applied masticatory forces are compressive in nature²¹.

Hence, the current study was conducted to assess the effect of essential oil modification of glass

ionomer cement on the antibacterial activity and compressive strength.

MATERIALS AND METHODS

Materials used in the study

In the current study, four commercially available essential oils were used; 2.5% bergamot, 2.5% peppermint, 2.5% lemongrass and 2.5 % fennel (Harraz for Food Industry and Natural Products, www.harraz.eg, Egypt). A conventional glass ionomer filling material was used in the current study (FILLBEST, BMS DENTAL S.r.l., Via M. Buonarroti, Zona Industriale 56033 Capannoli (PISA), WWW.bmsdental.it, Italy).

Analysis of the chemical components of the essential oils

Analysis of the chemical components of the essential oils used in the study was performed using gas chromatography–mass spectrometry analysis (GC-MS) at Central Laboratories Network, National Research Centre, Cairo, Egypt. The GC-MS system (Agilent Technologies) was equipped with gas chromatograph (7890B) and mass spectrometer detector (5977A). Essential oil samples were diluted using hexane (1:19, v/v). The GC was equipped with DB-WAX column (30 m x 250 µm internal diameter and 0.25 µm film thickness). Analysis was carried out using hydrogen as the carrier gas at a flow rate of 2.0 ml/min, injection volume of 1 µl and the following temperature program: 40 °C for 1 min; rising at 4 °C /min to 150 °C and held for 6 min; rising at 4 °C/min to 210 °C and held for 1 min. The injector and detector were held at 280 °C and 220 °C, respectively. Mass spectra were obtained by electron ionization (EI) at 70 eV; using a spectral range of m/z 50-550 and solvent delay 4 min. Identification of different constituents was determined by comparing the spectrum fragmentation pattern with those stored in Wiley and NIST Mass Spectral Library data ²².

Specimens grouping and preparation

A total of 100 specimens were prepared for the present study. Fifty specimens were prepared for the antibacterial activity test, 3 mm in diameter x 2 mm in height, and 50 specimens for the compressive strength test, 4 mm in diameter x 6 mm in height. Specimens for each test were prepared using a custom made split teflon mold with the intended dimensions. For each test, specimens were divided into five groups (n=10) according to the type of the essential oil used in modification of the glass ionomer cement; Group 1: unmodified glass ionomer cement (control group); Group 2: 2.5% bergamot modified GIC; Group 3: 2.5% peppermint modified GIC Group 4: 2.5% lemongrass modified GIC and Group 5: 2.5 % fennel modified GIC.

All specimens were freshly mixed with a powder / liquid ratio as recommended by the manufacturer. For unmodified glass ionomer specimens (control group), one scoop of powder and one drop of liquid were mixed on a clean glass slab until a homogenous mix was obtained. The mix was applied inside the split teflon mold which was placed on a glass slide covered with celluloid strip (Stripmat, POLYDENTIA, CH-6805 Mezzovico, Switzerland). Another celluloid strip was applied on the top surface of the split teflon mold and gently pressed under another glass slide to achieve well-packed smooth surfaced specimens. For essential oil modified glass ionomer cement specimens, one drop of the essential oil was applied to the drop of the glass ionomer liquid directly before manipulation, mixed with one scoop of powder and applied into the mold in the same manner as mentioned before.

Antibacterial activity test

Antibacterial activity was assessed using agar diffusion test. ATCC 25175 Type strain *Streptococcus mutans* (16S rRNA gene, Serotype c. carious dentin) were obtained from MIRCEN (Microbiological Resources Centre, Cairo, Egypt) to be used in this study. Bacteria were cultured

at 37°C overnight in Trypticase Soy agar (BD 236950, Difco TM, USA) and used as inoculums. The McFarland 0.5 turbidity standard (Densimat, BioMerieux, France) was used to determine the turbidity of the suspension. The concentration of bacteria is standardized to about 1xCFU/ml and used as a working microbial solution.

A total of 100 µl of the previously prepared working microbial solutions was spread evenly over Trypticase Soy agar. The plates were incubated at 37°C in a bacteriological incubator. The antibacterial effect for all experimental groups were assessed by measuring the diameter of bacterial growth inhibition zones after 24 and 48 h. The diameter of the bacterial growth inhibition zones was measured in millimeters using an electronic digital caliper at two different points to obtain two measurements for each specimen and then a mean value was determined for each one.

Compressive strength test

Specimens prepared for compressive strength testing were subjected to compressive loading using a universal testing machine (Lloyd LR 5K, Lloyd Instruments Ltd, Hampshire, UK) operating using Nexygen software version 4.6. The load was applied along the long axis of the specimens with a load cell 5KN at a cross head speed 0.5 mm/min. The maximum load at fracture was recorded and the compressive strength was calculated in MPa by dividing the load with the cross-sectional area of the specimen.

Statistical analysis

Numerical data were represented as mean and standard deviation (SD) values. Shapiro-Wilk's test was used to test for normality. Homogeneity of variances was tested using Levene's test. Data showed parametric distribution and variance homogeneity. Bacterial inhibition zone data were analyzed using two-way mixed model ANOVA.

Comparison of simple main effects was done utilizing one-way ANOVA followed by Tukey's post hoc test for independent variable and paired t-test for repeated measurements with p-values adjustment for multiple comparisons using Bonferroni correction. Compressive strength data were analyzed using one-way ANOVA followed by Tukey's post hoc test. The significance level was set at $p < 0.05$ within all tests. Statistical analysis was performed with R statistical analysis software version 4.1.3 for Windows*.

RESULTS

Results of chemical analysis of essential oils

Results of chemical analysis of different constituents of the essential oils used in the study are presented in tables 1-4.

Results of antibacterial activity (bacterial inhibition zone)

Bacterial inhibition zones as shown in table 5 revealed statistically significant differences between different essential oil modified GIC groups at both time intervals ($p < 0.001$). Bergamot recorded the highest mean value at both 24 h and 48 h, followed by fennel, then lemongrass and peppermint. The lowest mean value was revealed by the control group. Regarding time intervals, bergamot showed no significant difference between both time intervals ($p = 0.057$). However, other essential oils modified GIC groups reported significant increase of bacterial inhibition zones after 48 h ($p < 0.05$).

Results of compressive strength test

Results of compressive strength (table 6) presented a significant difference between different experimental groups ($p < 0.001$) with the control group showing the highest mean value, followed by lemongrass, then peppermint and bergamot. The lowest mean value was recorded by fennel modified GIC.

* R Core Team (2022). R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria. URL <https://www.R-project.org/>.

TABLE (1): Chemical components of bergamot essential oil

Peak	RT	Name	Formula	Area	Area Sum %
1	4.918	beta-Pinene	C10H16	1436508.16	5.77
2	6.159	beta-Myrcene	C10H16	522061.97	2.1
3	7.18	Limonene	C10H16	10672395.6	42.87
4	8.171	gamma-Terpinene	C10H16	1421897.98	5.71
5	8.337	Santolina triene	C10H16	145491.92	0.58
6	8.628	Prehnilol	C10H14	141376.17	0.57
7	16.955	trans-Geraniol	C10H18O	3711105.3	14.91
8	17.258	Linalyl acetate	C12H20O2	6707585.84	26.94
9	22.362	Linalyl propionate	C13H22O2	136275.26	0.55

Table (2): Chemical components of peppermint essential oil

Peak	RT	Name	Formula	Area	Area Sum %
1	4.847	beta-Pinene	C10H16	89321.03	0.56
2	6.871	Limonene	C10H16	275114.49	1.74
3	7.138	Eucalyptol	C10H18O	221432.6	1.4
4	8.568	Prehnilol	C10H14	62432.15	0.39
5	14.248	Cyclohexanone, 5-methyl-2-(1-methylethyl)-, cis-	C10H18O	3937906.12	24.87
6	14.937	1-Menthone	C10H18O	1320313.69	8.34
7	17.192	Linalyl acetate	C12H20O2	853735.12	5.39
8	18.13	neo-Menthol	C10H20O	874380.42	5.52
9	19.086	Pulegone	C10H16O	316184.04	2
10	19.566	Levomenthol	C10H20O	6131402.99	38.73
11	20.813	Camphene	C10H16	73264.36	0.46
12	21.478	Carvone	C10H14O	1675976.56	10.59

Table (3): Chemical components of lemongrass essential oil

Peak	RT	Name	Formula	Area	Area Sum %
1	6.123	beta-Myrcene	C10H16	316384.96	1.71
2	10.485	6-Methyl-5-hepten-2-one	C8H14O	184310.9	0.99
3	16.813	trans-Geraniol	C10H18O	176103.12	0.95
4	17.228	Linalyl acetate	C12H20O2	286783.22	1.55
5	20.445	Carveol	C10H16O	6743170.97	36.39
6	21.923	Z-Citral	C10H16O	9528558.03	51.42
7	22.439	Linalyl propionate	C13H22O2	496473.57	2.68
8	24.784	Geraniol formate	C11H18O2	799011.31	4.31

TABLE (4): Chemical components of fennel essential oil

Peak	RT	Name	Formula	Area	Area Sum %
1	6.865	Limonene	C10H16	477574.94	2.76
2	11.999	Fenchone	C10H16O	142100.55	0.82
3	19.941	Estragole	C10H12O	16004531.9	92.51
4	22.386	Linalyl propionate	C13H22O2	124798.78	0.72
5	23.532	2,4-Decadienal, (E,E)	C10H16O	181885.75	1.05
6	23.727	Anethole	C10H12O	368684.57	2.13

TABLE (5): Bacterial inhibition zone of different essential oil modified GIC groups at different time intervals

Time	Bacterial inhibition zone (mm) (Mean \pm SD)					f-value	p-value
	Control	Bergamot	Peppermint	Lemongrass	Fennel		
24h	0.00 \pm 0.00 ^E	8.51 \pm 0.17 ^A	6.19 \pm 0.03 ^D	7.19 \pm 0.06 ^C	7.58 \pm 0.03 ^B	8057.57	<0.001*
48h	0.00 \pm 0.00 ^E	8.73 \pm 0.08 ^A	6.25 \pm 0.06 ^D	7.32 \pm 0.04 ^C	7.64 \pm 0.04 ^B	22911.31	<0.001*
t-value	NA	7.03	10.86	26.00	32.35		
p-value	NA	0.057	0.030*	0.007*	0.005*		

NA: Not Applicable, means with different superscript letters within the same row are significantly different*significant

TABLE (6): Compressive strength of different essential oil modified glass ionomer cements

Compressive Strength (MPa) (Mean \pm SD)					f-value	p-value
Control	Bergamot	Peppermint	Lemongrass	Fennel		
131.83 \pm 3.01 ^A	84.55 \pm 4.61 ^D	105.28 \pm 7.91 ^C	116.20 \pm 5.61 ^B	73.04 \pm 3.09 ^E	104.79	<0.001*

Means with different superscript letters within the same row are significantly different*significant (p<0.05)

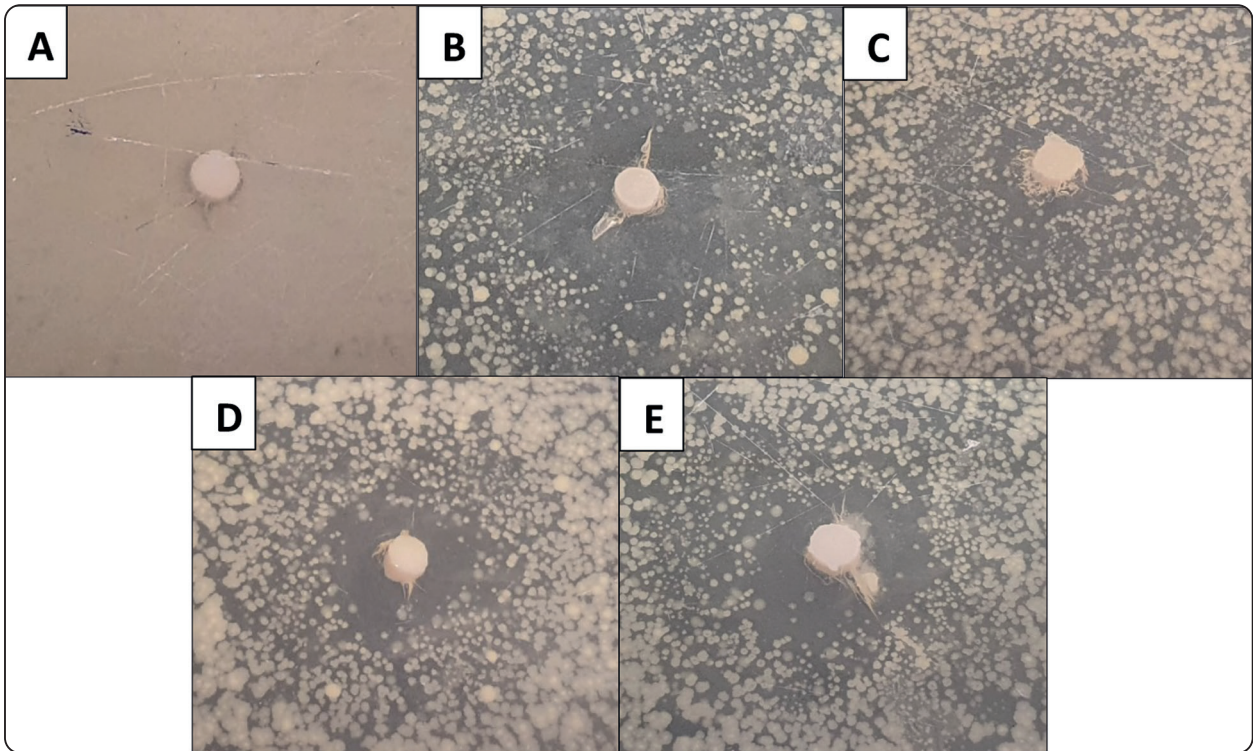


Fig. (1): Bacterial inhibition zone of all experimental groups; (A): Unmodified GIC (Control), (B): Bergamot modified GIC, (C): Peppermint modified GIC, (D): Lemongrass modified GIC and (E): Fennel modified GIC

DISCUSSION

The current study assessed the effect of essential oil modification of glass ionomer cement on antibacterial activity and compressive strength. Medicinal plants are considered major source of a wide variety of chemical compounds that possess multiple biological activities as antifungal and antibacterial activities in addition to treatment of other various diseases¹³.

Results of the current study revealed that all tested essential oil modified GIC groups showed significantly higher antibacterial activity in comparison to the control group. Essential oils possess multiple antibacterial mechanisms. They can attack the cytoplasmic membrane of bacterial cell increasing its permeability which subsequently affects its electron transport function, nutrient absorption, nucleic acid synthesis and ATPase enzyme activity^{23,24}. Hydrophobicity of essential oils allows them to penetrate bacterial membrane

phospholipids and mitochondria leading to bacterial cell death¹⁶.

In addition, phenolic compounds found in essential oils enhance their antimicrobial properties specially against gram positive bacteria²⁵. Interaction of the hydroxyl group of phenolic compounds with the bacterial cytoplasmic membrane changes its permeability and affects lipid ordering and stability leading to disruption of the cellular components²⁶.

Bergamot modified GIC showed the highest bacterial inhibition zone than the control group and other essential oil modified GIC groups (table 5). This might be attributed to its chemical composition. The monoterpene limonene is the major component in citrus essential oils ranging from 32 to 98%²⁷. According to the results of the gas chromatography–mass spectrometry analysis of the current study (table 1), limonene recorded the highest percentage of the constituents of bergamot representing 42.87%. *Subramenium et al, 2015*²⁸,

found that limonene has 75-95 % antibiofilm activity against streptococcus mutans. They stated that limonene is able to inhibit bacterial adhesion to tooth surface and inhibit bacterial acid production in addition to its ability to downregulate several genes (covR, mga and vicR). Those genes play a critical role in regulating surface-associated proteins in streptococcus mutans.

Presence of other constituents as linalyl acetate and trans-Geraniol could potentiates its antibacterial activity²⁹. Bergamot essential oil used in the current study is composed of linalyl acetate (26.94 %) and trans-Geraniol (14.91 %) as reported in table 1. Linalyl acetate is able to disrupt the lipid fraction of microorganism plasma membrane, resulting in alterations of cell membrane permeability and leakage of intracellular materials³⁰. *Guimarães et al, 2019*³¹, reported that alcohol compounds like Trans-geraniol is one of the effective and fast acting antimicrobial agents and attributed this effect to the presence of hydroxyl groups. Another antibacterial mechanism of geraniol was described via its ability to adhere to bacterial cell membrane lipids and interacts with its components and hence increase its permeability in addition to its ability to bind to important intracellular sites resulting in cell destruction³².

The higher percentage of the previously mentioned constituents of bergamot essential oil in addition to their antimicrobial properties via multiple mechanisms might explain its highest antibacterial results in comparison to other experimental groups.

According to the findings of the current study, peppermint recorded antibacterial activity higher than the control group, however it was the least among all tested essential oil groups. *Sreevidhya, 2014*³³, revealed an antibacterial activity of peppermint against streptococcus mutans. This finding was previously explained by the presence of menthol and carvone constituents¹⁶. Also, *Ambrosio et al, 2021*³⁴, reported antibacterial potential and

antioxidant activity for essential oils containing menthol and carvone.

Peppermint essential oil used in the current study is composed of 38.73 % Levomenthol, 5-52% neo-menthol and 8.34% I-menthone (table 2). Menthol showed an antibacterial mechanism as that described before for linalyl acetate monoterpene based on its lipophilicity which make it able to disrupt the lipids of microorganism plasma membrane with negative impact on cell permeability³⁰. Carvone represents 10.59 % of the chemical composition of peppermint (table 2). It is considered a powerful antimicrobial agent able to penetrate bacterial cell membrane increasing cell permeability³⁵. In addition to the presence of minor constituents as linalyl acetate (5.39 %) and limonene (1.74 %).

Lemongrass essential oil showed antibacterial activity higher than the control group which might be attributed to its chemical composition (table 3). *Ilango et al, 2019*⁹, reported antibacterial activity of lemongrass essential oil against streptococcus mutans at minimum inhibitory concentration of 10 μ l. The major component of lemongrass as reported in table 3 is Z-Citral which represents more than 50% of the components. Z-citral is a monoterpene which causes loss of membrane integrity of bacterial cells increasing cell membrane permeability via hydrophobic interactions with the membrane^{36,37}. It could also induce changes in the intracellular pH and intracellular ATP concentration in the bacterial cell as reported by *De Silva et al, 2017*³⁸.

Presence of carveol as one of the major components of lemongrass (36.39%) plays a great role in its antibacterial activity. It has been previously reported that carveol has a strong antibacterial activity against gram positive bacteria^{31,34}. *Lopez-Romero et al, 2015*³⁹, recorded changes in the hydrophobicity, surface charge and bacterial cell membrane integrity after exposure to essential oils containing carveol.

Fennel reported antibacterial activity against

streptococcus mutans higher than control group, lemongrass and peppermint essential oil modified groups. Previous research demonstrated an antibacterial activity of fennel against streptococcus mutans¹⁶. According to the chemical analysis results of the current study (table 4), the major constituent of fennel EO is estragole representing 92.51 %. *Andrade et al, 2015*⁴⁰, reported high inhibition zone of estragole against gram positive bacteria.

Several mechanisms could be responsible for the development of microbial resistance to antibacterial agents as poor diffusion of the antibiotics through the biofilm, phenotypic changes of the cells forming the biofilm and the expression of efflux pumps. Efflux pumps allow the microorganisms to regulate their internal environments by removing toxic substances such as antimicrobial agents and metabolites⁴¹. Hence, efflux pump inhibitors are important to enhance the efficiency of antimicrobial agents. *Da Costa et al, 2021*⁴², demonstrated that estragole is a potent efflux pump inhibitor.

It has been reported that the biological activity of the major components of essential oils is modulated by the minor components. The synergism among the major and minor components potentiates the antimicrobial activity of the essential oils^{34,43}. *Alexa et al, 2019*¹⁴, highlighted the effect of synergism of different components by testing antibacterial activity of different essential oils in binary and tertiary mixtures where they reported higher results than using essential oil in a single form. The findings of the current study cannot be directly correlated with other studies since, no previous researches investigated the effect of essential oil modification of GIC using bergamot, peppermint, lemongrass and fennel on the antibacterial activity.

Though the tested essential oils improved the antibacterial potential of glass ionomer cement, it is important not to jeopardize its mechanical properties. Compressive strength results demonstrated a significant decrease for all essential oil modified

GIC groups in comparison to the control group. *Sherief et al, 2021*⁴⁴, reported low compressive strength of glass ionomer cement modified using cinnamon and thyme essential oil. They explained this finding by the disability of essential oils to chemically bond to the glass and the polyalkenoate matrix resulting in high structural micro-porosity and diminished strength properties. Another explanation was attributed to the fact that cross-linking in GIC occurred due to coordination of Al^{3+} and Ca^{2+} with the COOH groups of the polyacrylic acid. Presence of essential oil within GIC deprives many COOH groups from interlinking within the glass ionomer matrix and causes deceleration of the setting reaction which adversely affect the compressive strength^{10, 44}. In addition, changing the powder/liquid ratio of GIC by adding essential oil to polyacrylic acid during material manipulation could adversely affect its mechanical properties¹⁰.

Among essential oil modified groups, lemongrass recorded the highest compressive strength value followed by peppermint then bergamot and the least value was demonstrated by fennel. This might be attributed to chemical structure of different components of the tested essential oils. Compounds containing functional OH and COOH groups as those present in the chemical structure of glass ionomer for inducing acid-base reaction between glass particles and polyacrylic acid might improve compressive strength of GIC⁴⁵. Compressive strength results (table 6) demonstrated that lemongrass (116.20 ± 5.61) and peppermint (105.28 ± 7.91) essential oils modified GIC groups recorded mean values above 100 MPa. Although these values were significantly lower than the control group, it satisfies the requirement of the ISO standards 9917-1:2007 which reported that compressive strength of glass ionomer cements should not be less than 100 MPa⁴⁶.

Among essential oil modified groups, bergamot and fennel recorded higher antibacterial activity in

comparison to lemongrass and peppermint while, for compressive strength results, lemongrass and peppermint showed higher results. It seems like there is an inverse relation between antibacterial activity and compressive strength. In the agar diffusion test, the diameter of inhibition zone is greatly affected by the solubility and diffusion characteristics of the substances being tested⁴⁷. Although solubility of glass ionomer and rate of diffusion of the essential oil might positively affect the antibacterial activity, they also negatively affect the compressive strength.

CONCLUSIONS

Within the limitations of the current study, it could be concluded that essential oil modification of GIC was effective in enhancing antibacterial activity although it adversely affects compressive strength in comparison to the control group. Lemongrass and peppermint essential oil modification of GIC was effective in improving antibacterial property while preserving an acceptable compressive strength according to the ISO standards. Nevertheless, further studies should be conducted to assess the effect of the tested essential oils in different concentrations on other mechanical, physical and optical properties.

REFERNCES

1. Tatekalva P, Subbaiyan H and Kumar SR. Comparative evaluation of antimicrobial potential of herbal extracts on *Streptococcus mutans* and *Enterococcus faecalis* An in vitro study. *Braz Dent Sci* 2021; 24(1).
2. Wiwattanarattanabut K, Choonharuandej S and Srithavaj T. In Vitro anti-cariogenic plaque effects of essential oils extracted from culinary herbs. *J Clin Diagn Res* 2017; 11(9): DC30-DC35.
3. Loe H. Oral hygiene in the prevention of caries and periodontal disease. *Int Dent J* 2000; 50(3): 129-39.
4. Oyanagi T, Tagami J and Matin K. Potentials of mouthwashes in disinfecting cariogenic bacteria and biofilms leading to inhibition of caries. *Open Dent J* 2012; 6: 23-30.
5. Nakajo K, Imazato S, Takahashi Y, Kiba W, Ebisu S and Takahashi N. Fluoride released from glass-ionomer cement is responsible to inhibit the acid production of caries-related oral streptococci. *Dent Mater* 2009; 25: 703-708.
6. Prasad MP and Maradia MA. Antibacterial activity of conventional and modified glass ionomer cement against *Streptococcus mutans*. *J Appl Biol Biotech* 2014; 2(03): 017-020.
7. Sultan MS. Antibacterial effect of aloe vera and glass ionomer modified by aloe vera on streptococcus mutans. 2019; 65(3): 2607-2616.
8. Castilho ARF, Duque C, Kreling PF, Pereira JA, de Paula AB, Sinhoreti MAC et al. Doxycycline-containing glass ionomer cement for arresting residual caries: an in vitro study and a pilot trial. *J Appl Oral Sci* 2018; 26: e20170116.
9. Ilango P, Suresh V, Vummidi AV, Ravel V, Chandran V, Mahalingam A et al. Evaluation of antibacterial activity of lemongrass oil against oral clinical isolates – An in vitro study. *Pharmacogn J* 2019; 11(5):1023-1028.
10. Mittal S, Soni H, Aharma DK, Mittal K, Pathania V and Sharma S. Comparative evaluation of the antibacterial and physical properties of conventional glass ionomer cement containing chlorhexidine and antibiotics. *J Int Soc Prev Community Dent* 2015; 5(4): 268- 275.
11. Seow YX, Yeo CR, Chung HL and Yuk H-G. Plant essential oils as active antimicrobial agents. *Crit Rev Food Sci Nutr* 2014; 54: 625-644.
12. Chouhan S, Sharma K and Guleria S. Antimicrobial activity of some essential oils-Present status and future perspectives. *Med* 2017; 4: 58.
13. Rêgo CB, Silva AM, Gonçalves LM and Paschoal MAB. In vitro antimicrobial activity of essential oil of *Cymbopogon citratus* (lemongrass) on *Streptococcus mutans* biofilm. *Afr J Microbio Res* 2016; 10(31): 1224-1228.
14. Alexa VT, Galuscan A, Popescu I, Tirziu E, Obistioiu D, Floare AD et al. Synergistic/Antagonistic potential of natural preparations based on essential oils against streptococcus mutans from the oral cavity. *Molecules* 2019; 24: 4043.
15. Navarra M, Mannucci C, Delbò M and Calapai G. Citrus bergamia essential oil: From basic research to clinical application. *Front Pharmacol* 2015; 6: 36.
16. Golestannejad Z, Gavanji S, Mohammadi E, Motamedi A, Bahrani M, Rezaei F et al. Comparison of antibacterial

- activity of essential oils of *Foeniculum vulgare* Mill, *Mentha arvensis* and *Mentha piperita* against *Streptococcus mutans*. *Adv Herb Med.* 2017; 3(1): 3-13.
17. Shah G, Shri R, Pancha IV, Sharma N, Singh B and Mann AS. Scientific basis for the therapeutic use of *Cymbopogon citratus*, Stapf (Lemongrass). *J Adv Pharm Technol Res* 2011; 2(1): 3-8.
 18. Gulfracz M, Mehmood S, Minhas N, Jabeen N, Kausar R, Jabeen K et al. Composition and antimicrobial properties of essential oil of *Foeniculum vulgare*. *Afr J Biotechnol* 2008; 7(24): 4364-4368.
 19. Enezes-Silva R, de Oliveira BMB, Magalhaes APR, Bueno LS, Borges AFS, Baesso ML et al. Correlation between mechanical properties and stabilization time of chemical bonds in glass-ionomer cements. *Braz Oral Res* 2020; 34: e053.
 20. Hafez RM, Elkorashy ME and Sultan MS. Compressive strength and surface roughness of ceramic reinforced glass ionomer subjected to chemical challenge. *Egy Dent J* 2017; 63: 2709-2719.
 21. Enezes-Silva R, Cabral RN, Pascotto RC, Borges AFS, Martins CC, Navarro MFD et al. Mechanical and optical properties of conventional restorative glass-ionomer cements – a systemic review. *J App Oral Sci* 2019; 27: e2018357.
 22. Wiley Registry 11th Edition/NIST 2017 Mass Spectral Library.
 23. Dinesh M, Uma M, Anjali V, Neetushree M and Shanmugam V. Inhibitory properties of aqueous extracts of selected indigenous medicinal plants against dental caries causing *Streptococcus mutans* and *Streptococcus mitis*. *Afr J Basic Appl Sci* 2013; 5(1): 8-11.
 24. Zaker SR, Gavanji S, Sayedipour SS, Bakhtari A, Bidabadi EH, Larki B et al. The effect of some herbal essential oils on pathogenic bacteria. *Ethno Pharma Pro* 2015; 1(2): 23-34.
 25. Lourenço M, Bernardi ACA, Lunardi N, Neto RJB, Bernardi PSM and Boeck EM. In vitro evaluation of the antimicrobial activity of Basil (*Ocimum basilicum* L.) and Coriander (*Coriandrum sativum* L.) oil extracts on *Streptococcus mutans*. *J Res Dent* 2017; 5(2): 40-45.
 26. Ahmad A, Vuuren SV and Viljoen A. Unravelling the complex antimicrobial interactions of essential oils-the case of *Thymus vulgaris* (thyme). *Molecules* 2014; 19: 2896-2910.
 27. Mahato N, Sharma K, Koteswararao R, Sinha M, Baral E and Cho MH. Citrus essential oils: Extraction, authentication and application in food preservation. *Crit Rev Food Sci Nutr* 2019; 59: 611- 625.
 28. Subramenium GA, Vijayakumar K and Pandian SK. Limonene inhibits streptococcal biofilm formation by targeting surface-associated virulence factors. *J Med Microbiol* 2015; 64: 879-890.
 29. Lapinska B, Szram A, Zarzycka B, Grzegorzczak J, Hardan L, Sokolowski J et al. An in vitro study on the antimicrobial properties of essential oil modified resin composite against oral pathogens. *Mater* 2020; 13: 4383.
 30. Trombetta D, Castelli F, Sarpietro MG, Venuti V, Cristani M, Daniele C et al. Mechanisms of antibacterial action of three monoterpenes. *Antimicrob agents Chemother* 2005; 49(6): 2474- 2478.
 31. Guimarães AC, Meireles LM, Lemos MF, Guimarães MCC, Endringer DC, Fronza M et al. Antibacterial activity of terpenes and terpenoids present in essential oils. *Molecules* 2019; 24: 2471.
 32. De Lira MHP, Júnior FPD, Moraes GFQ, Macena GD, Pereira FD and Lima IO. Antimicrobial activity of geraniol: an integrative review. *J Essent Oil Res* 2020; 32(11050): 1-11.
 33. Sreevidhya TM. Antibacterial activity of the three essential oils on *Streptococcus mutans*- an in-vitro study. *Int J Drug Dev Res* 2014; 6 (4): 65-67.
 34. Ambrosio CMS, Diaz-Arenas GL, Agudelo LPA, Stashenko E, Contreras-Castillo CJ and Da Gloria EM. Chemical composition and antibacterial and antioxidant activity of a citrus essential oil and its fractions. *Molecules* 2021; 26: 2888.
 35. Bouyahya A, Mechchate H, Benali T, Ghchime R, Charfi S, Balahbib A et al. Health benefits and pharmacological properties of carvone. *Biomolecules* 2021; 11: 1803.
 36. Oliveira MMM, Brugnera DF, Cardoso MG and Piccoli RH. Yield, chemical composition and antilisterial composition of essential oils of *Cymbopogon* species. *Rev Bras Plantas Med.* 2011; 12: 8-15.
 37. Jiamboonsri P and Kanchanadumkerng P. Influence of gallic acid and thai culinary essential oils on antibacterial activity of nisin against *streptococcus mutans*. *Adv Pharmacol Pharmaceut Sci* 2021; Article ID 5539459: 12 p.

38. De Silva BCJ, Jung W, Hossain S, Wimalasena SHMP, Pathirana HNKS and Heo G. Antimicrobial property of lemongrass (*Cymbopogon citratus*) oil against pathogenic bacteria isolated from pet turtles. *Lab Anim Res* 2017; 33(2): 84-91.
39. Lopez-Romero JC, González-Ríos H, Borges A and Simões M. Antibacterial effects and mode of action of selected essential oils components against *Escherichia coli* and *Staphylococcus aureus*. *Evid -based Complement Altern Med* 2015; 2015: ID 795435.
40. Andrade TCB, De Lima SG, Freitas RM, Rocha MS, Islam T, Da Silva TG et al. Isolation, characterization and evaluation of antimicrobial and cytotoxic activity of estragole, obtained from the essential oil of *Croton zehntneri* (Euphorbiaceae). *Ann Braz Acad Sci* 2015; 87(1): 173-182.
41. Liu J, Zhang J, Guo L, Zhao W, Hu X and Wei X. Inactivation of a putative efflux pump (LmrB) in *Streptococcus mutans* results in altered biofilm structure and increased exopolysaccharide synthesis: implications for biofilm resistance. *Biofouling* 2017; 33(6): 481-493.
42. Da Costa RHS, Rocha JE, De Freitas TS, Pereira RLS, Junior FNP et al. Evaluation of antibacterial activity and reversal of the NorA and MepA efflux pump of estragole against *Staphylococcus aureus* bacteria. *Arch Microbiol* 2021; 203: 3551-3555.
43. Delamare APL, Moschen-Pistorello IT, Artico L, Atti-Serafini L and Echeverrigaray S. Antibacterial activity of essential oils of *Salvia officinalis* L. and *Salvia triloba* L. cultivated in South Brazil. *Food Chem* 2007; 100: 603-608.
44. Sherief DI, Fathi MS and Abou ElFadl RK. Antimicrobial properties, compressive strength and fluoride release capacity of essential oil-modified glass ionomer cements-an in vitro study. *Clin Oral Invest* 2021; 25: 1879-1888.
45. Frouzanmehr M, Barekatalin M and Shahtalebi MA. An in vitro evaluation of the compressive strength and shear bond strength of resin-modified glass-ionomer cement containing purified powder of *Salvia officinalis*. *Dent Res J* 2020; 17 (5): 319- 325.
46. International Organization for Standardization – ISO. ISO 9917-1:2007 – Dentistry: Water based cements. Part 1: Powder/liquid acid-based cements. Geneva: International Organization for Standardization; 2007.
47. Chaudhari LKD, Jawale BA, Sharma S, Kumar HSCDM and Kulkarni PA. Antimicrobial activity of commercially available essential oils against *Streptococcus mutans*. *J Contemp Dent Pract* 2012; 13(1): 71-74.