

Vol. 71, 303:315, January, 2025

PRINT ISSN 0070-9484 • ONLINE ISSN 2090-2360



Oral Medicine, X-Ray, Oral Biology and Oral Pathology

 Accept Date: 24-12-2024 Available online: 10-01-2025 • DOI: 10.21608/edj.2024.327835.3230

ALVEOLAR RIDGE PRESERVATION COMPARING ALLOGRAFT, ALLOPLAST, AND AUTOGENOUS TOOTH GRAFT: A RANDOMIZED CLINICAL TRIAL

Hadeel Gamal* , Yasmine Ahmed Fouad** , Basma Abdelrahman Ahmed***

and Susan Sarhan****

ABSTRACT

Introduction: Following tooth extraction, sequels of events that reduce alveolar crest width and height take place. Alveolar ridge preservation (ARP) after extraction of a tooth is crucial as it maintains ridge dimensions facilitating dental implant placement. Previous systematic reviews and clinical trials recommended allografts, alloplasts, and autogenous tooth bone graft (Auto-BT). However, the results were heterogeneous about the exact superiority of one material over the other. Thus, the herein work aimed to evaluate the efficiency of the different bone graft materials in ARP.

Material and Methods: In this randomized clinical trial, thirty patients were divided into three groups. ARP was performed using allograft, alloplast, and Auto-BT. Changes in alveolar ridge dimensions were evaluated clinically, radiographically, and histomorphometrically after 3 months.

Results: Results of intergroup comparisons for clinical and radiographic measurements showed higher bone loss values in the allograft and alloplast groups than in the Auto-BT group. Histological measurements disclosed that the Auto-BT group had the highest value of mature bone followed by alloplast while allograft presented the lowest value.

Conclusion: In conclusion, tooth graft could be considered a viable alternative to other graft materials in ARP. Autogenous tooth graft is the new concept of graft material. They can be processed and used as an economical, natural, and biocompatible, versatile, and predictable grafting material.

KEYWORDS Alveolar ridge augmentation; Allogenic graft; Alloplastic graft; Tooth graft

^{***} Assistant Professor of Oral Medicine and Periodontology, Faculty of Dentistry, Misr International University *** Lecturer of Oral Pathology, Faculty of Dentistry, Ain Shams University



^{*} Lecturer of Oral Medicine and Periodontology, Faculty of Dentistry, Ain Shams University

^{**} Assistant Professor of Oral Medicine and Periodontology, Faculty of Dentistry, Ain Shams University

(304) E.D.J. Vol. 71, No. 1 Hadeel Gamal, et al.

INTRODUCTION

Following tooth extraction, sequence of events eventually occur altering alveolar ridge dimensions and subsequently reducing alveolar crest width and height. The first 6 months after tooth extraction experience the most important changes with an average vertical and horizontal bone resorption of 1.24 mm and 3.79 mm respectively [1,2].

A range of alveolar ridge preservation (ARP) techniques has been suggested to compensate for the variations previously mentioned. Generally, the foundation of these techniques comprises emplacement of bone graft into the tooth socket, instantly after extraction and sealing the socket with a barrier. Evaluation of the efficacy of these interventions has been well studied and, the efficiency of ARP procedures in bone resorption reduction compared to treatment without ARP is well reported [3,4]

Alveolar ridge preservation after tooth extraction is crucial for dental implant success as it maintains dimensions that facilitate dental implant placement. One another goal for ARP is to afford vital osseous tissue at the extracted tooth site, which will eventually hold up the implant and provide osseointegration. Diverse materials and techniques are used to achieve these two goals comprising autogenous tissues, allografts, alloplastics, and xenografts [4-6].

Guided bone regeneration (GBR) aims at preventing gingival epithelial cells and connective tissue from entering the socket by cell occlusive membranes. Resorbable and non-resorbable barrier membranes are sometimes used to keep space for bone formation and growth. Having an advantage over non-resorbable membranes, resorbable membranes show good healing of soft tissues and do not need a second surgery to remove [7]. In addition to the profits of collagen in helping clot organization and stabilization, collagen membranes could be effortlessly manipulated, and adapted to bone [8].

Allograft materials from cadavers are usually obtained through tissue banks. They have both osteoinductive and osteoconductive properties [9]. Having a slower resorption rate, allografts can keep the ridge dimensions stable [2,4]. Small allograft particles may remain up to a year before complete resorption. Creeping substitution at the recipient place and connective tissue is the method of revascularization of freeze-dried bone allografts (FDBA) [10]. The quantity of the newly formed bone and the dimensional stability of both cortical and cancellous FDBA were similar when used in ARP [11].

Alloplastics such as beta tricalcium phosphate (β -TCP) represent a group of largely available synthetic bone substitutes. They are osteoconductive, biocompatible, and do not carry any risk of infection or disease transmission [12, 13]. They act as biologic fillers with restricted periodontal regeneration [10,14].

Recently, autogenous tooth bone graft material (Auto-BT) has gained much attention in dental implant augmentation through GBR ^[15]. The production of Auto-BT from the teeth after extraction has been approached in a number of ways. The fillings, soft tissues, and carious parts should be removed following tooth extraction ^[16]. Some protocols advise using the extracted tooth's root alone ^[17], while others support using both the crown and root ^[18]. Some research has confirmed the excellent clinical and histological outcomes of the deciduous and permanent tooth- based Auto-BT graft materials ^[19,20].

The superiority of Auto-BT lies in its resemblance to autogenous bone in both its histological structure and components [21,22]. The composition of both human dentin and bone is 65% inorganic and 35% organic [23]. Auto-BT ensures excellent biocompatibility with no fear of immune rejection. In addition, many studies have reported that Auto-BT exhibits not only osteoconductive but also osteoinductive capability [20, 24, 25]. The osteoconductivity is linked to the inorganic proportions [26] while the organic

matrix of mineralized dentin is responsible for its osteoinductive characteristics ^[27, 28]. The hydroxyapatite in dentin is in the form of calcium phosphate with low crystal content making it easily degradable by the osteoclastic activity. However, the organic content consists of type I collagen network (90%), non-collagenous proteins (10%) (osteocalcin, osteonectin, sialoprotein, and phosphoprotein) which aid in calcification of bone, and some growth factors (bone morphogenetic proteins, and insulin-like growth factor) which give the tooth its osteoinductive properties ^[29].

Due to the paucity of studies specifying the superiority of one material over the other in ARP, the present study aimed to compare the efficiency of allograft, alloplast and Auto-BT graft materials on ARP subsequent to tooth extraction.

MATERIAL AND METHODS

This study received an approval from the ethics committee of the Faculty of Dentistry, Ain Shams University, Cairo, Egypt (IRB no: FDASU-Rec IR092210) and registered in Clinical Trials (ID: NCT05812872). It was performed according to the Declaration of Helsinki

Thirty patients were selected from the outpatient clinic of Oral Medicine, Periodontology, and Oral Diagnosis department, Faculty of Dentistry, Ain Shams University. A power analysis was designed to have adequate power to apply a statistical test of the null hypothesis that there is no difference would be detected between tested groups. By adopting an alpha (α) level of (0.05), a beta (β) level of (0.2) (i.e. power=80%), and an effect size (f) of (0.637) calculated based on the results of a previous study ^[30]; the minimal required sample size (n) was found to be (27) cases (i.e. 9 cases per group). Sample size calculation was performed using G*Power version 3.1.9.7 ^[31].

Inclusion Criteria

- a) Healthy adult patients as evidenced by Burket's oral medicine health history questionnaire [32].
- b) Both genders
- c) Age ranging from 20 40 years old
- d) Having at least one hopeless tooth (traumatized, badly broken, unrestorable, etc.) indicated for extraction upper / lower (Anterior or premolar area)
- e) Sockets type I or II.

Consent was written by patients after explaining the nature of the study.

Exclusion Criteria

- a) Smokers
- b) Pregnant and breast-feeding females
- c) Mentally retarded patients
- d) Handicapped patients and prisoners
- e) Teeth with periodontal or periapical infections
- f) Patients with malocclusion
- g) Patients with parafunctional habits
- h) Patients receiving drugs that may influence bone metabolism

Study Design and Patient Grouping

This study is a clinical comparative prospective study. Patients who met the eligibility criteria were allocated into: allograft group, alloplast group and autogenous tooth graft group.

The randomization of patients into the three groups was performed by a computer-generated randomization list. Allocation blindness was attained by putting the subject's treatment in a sealed envelope. The patients, outcome examiners and the statistician were blinded.

Each group included 10 patients who had undergone single tooth extraction, then the socket

(306) E.D.J. Vol. 71, No. 1 Hadeel Gamal, et al.

was filled by either mineralized cortico-cancellous allograft (Maxgraft, botiss dental GmbH, Berlin–Germany), β -TCP bone graft (Bioresorb, Implant direct, CA, USA) or Auto-BT. All three materials in each group were loaded with collagen membrane (Hyprosorb-F Atelo collagen membrane).

Surgical Procedure

Extraction and Socket Augmentation Procedure (Fig.1)

- a) Baseline cone beam computed tomography (CBCT) was taken on the extraction day for socket type evaluation.
- b) Patients were injected with anesthesia (Artinibsa 40mg/0.01 mg/ml solution injectable-inisba-Spain), extraction was performed atraumatically, using periotome (Nordent Germany), to preserve bone and soft tissue, then extraction was completed using forceps (Martin-Nelson-Germany).
- c) Curettage of socket was carried out by bone curettes (Reicodent-Germany).

d) Clinical measurements of bone height and width were performed as follows:

Bone height

Bone height was assessed using a periodontal probe (Hu-Friedy UNC 15 Co., LLC-USA) till reaching the bone. A stent of thickness one mm was fabricated before extraction using a cast. The tooth to be extracted was removed from the cast [33]. Six holes were made in the resin plate in the following positions: mesio buccal, mid buccal, disto buccal, mesio palatal, mid palatal, and disto palatal. Measurements were taken after tooth extraction (baseline) and 3 months after extraction before placement of the implant.

Rone Width

Alveolar ridge width was measured immediately after extraction and after 3 months using a caliper clamp (Lwebinger GmbH.Mulheim, Germany), the width was measured perpendicular to the tangent of the dental arch at the mid-point of the extraction site approximately 4 mm apical to the marginal gingiva of the adjacent teeth [34].

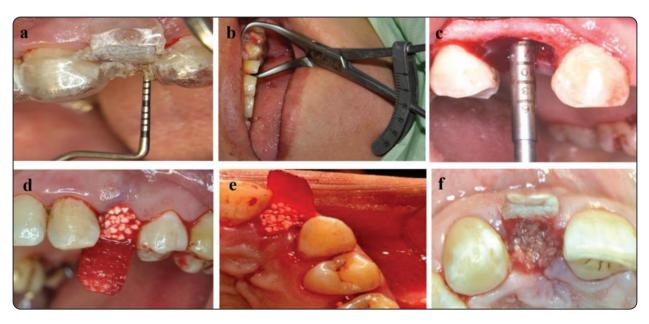


Fig. (1) Photomicrographs showing: (a) Periodontal probe measuring mesiobuccal bone height, (b) Measuring bone width immediately after extraction using a bone caliper. (c) Socket filled with mineralized allograft, (d) Socket filled with beta tricalcium phosphate alloplast, (e) Socket filled with autogenous tooth graft.

- e) Sockets were filled with mineralized corticocancellous allograft, β -TCP bone graft (500-1000 μ m), and Auto-BT.
- f) All sockets were covered by collagen membrane to cover the graft and stabilize the blood clot.
- g) Socket approximation was performed using 5/0 reverse cutting 3/8th vicryl suture.
- h) Amoxicillin (Amoxil-GlaxoSmithKline, medical union pharmaceuticals, Egypt) 500 mg t.d.s, Metronidazole Flagyl, Sanofi Aventis, Egypt) 500 mg twice/day and antiseptic (Hexitol, Arab drug company, Egypt) mouth wash for 1 week) were prescribed.
- Post-operative instructions were given to the patients; the patients were instructed not to wear any prosthetic restoration.

Preparation of Autogenous Tooth Graft

Method: from extraction to grafting particulate dentin

Extraction of teeth without any root canal restorations was done and prepared for immediate grafting.

- a) Crowns or restorations were removed. Caries, areas of dentin discoloration, calculus and periodontal ligament (PDL) remnants were reduced, and multi-rooted teeth were split.
- b) Air syringe was used to dry clean teeth and grinded into the grinding sterile chamber of the Smart Dentin Grinder (SDG) (Kometa Bio ltd., London, United Kingdom)^[35].
- c) The roots were ground in 3 seconds by the SDG. The vibrating movement of the grinding chamber then filtered and collected particles between 300µm and 1,200µm into a lower chamber. Smaller particles usually fell into a waste drawer, as this fine particulate is incompatible for bone grafting. This protocol was repeated to grind the remaining teeth particles. The collected particulate dentin was immersed in basic alcohol (0.5M of NaOH and 30% alcohol (v/v)) for 10 minutes for dissolving any fats,

organic debris or bacteria. The particulate was then washed twice in sterile phosphate-buffered saline (PBS) leaving wet particulate dentin that was grafted into the extracted sockets.

Implant Placement and Core Biopsy Procedure

- a) After 3 months, another CBCT was done. Changes in the width and height measurements at the center of the extraction socket were evaluated in merged axial and sagittal views using the Romexis superimposition system besides measuring the changes in density (**Fig.2**).
- b) Before implant placement, clinical measurements of height and width were repeated.
- c) Reflection of an open flap was done for taking a core biopsy using trephine bur (Hu-Friedy trephine bur TREO20), and placement of a submerged implant (SIC Invevt AG Birmannsgasse 3 CH-4055 Basel, Germany) and then flaps were closed.

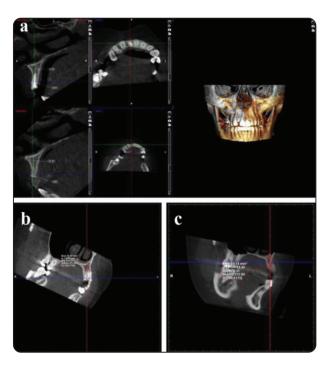


Fig. (2) Photomicrographs showing: (a) superimposition of CBCT base line and after three months to detect bone height and width, (b,c) measurements of preoperative and postoperative bone density, respectively.

(308) E.D.J. Vol. 71, No. 1 Hadeel Gamal, et al.

Histological Examination and Histomorphometric Analysis

The taken bone specimens were fixed in 10% buffered formalin for five days. The specimens were decalcified using a solution containing 12% Ethylene diamine tetra-acetic acid (EDTA) buffered in pH 7.2 PBS for three weeks at 4°C ^[36], then the specimens were assigned for staining and histomorphometric analysis. Specimens were infiltrated and embedded in the center of paraffin wax blocks after being washed properly under running water, dehydrated by ascending concentrations of alcohol, and transferred to xylol. The embedded specimens were sectioned by microtome (4 microns thick) and stained by hematoxylin and eosin (H &E) and Masson trichrome (MT) stains.

Representative photomicrographs of H &E sections and three microscopic fields of each MT stained section were captured at a magnification of 200X using a digital camera (Canon EOS

650D) mounted on a light microscope (BX60, Olympus, Japan). H &E sections were used for histological evaluation while MT special stain was used to detect areas of immature and mature collagen. The immature collagen appeared blue while areas with mature collagen appeared reddish [37]. The MT images were analyzed to obtain the area percentage of mature and immature collagen of the newly formed bone besides the area percentage of the residual graft materials using image J (1.41a, NIH, USA) software.

Statistical analysis

R statistical analysis software version 4.1.3 for Windows (R Core Team 2022) was used for the statistical analysis. Mean and standard deviation (SD) values of numerical data were calculated. Shapiro- Wilk's test was performed to test for normality. Data were normally distributed and were analyzed using one-way ANOVA followed by Tukey's post hoc test. Unpaired T test was used when comparing 2 groups. The significance level was set at p<0.05 within all tests.

RESULTS

Thirty patients were classified equally and randomly into three groups, each group included 10 patients. The allograft group included six males and four females with a mean age of 31.4 years (range 24-40 years). The alloplast group included (four males and six females with a mean age of 29.4 years (range 21-38 years). While, the Auto-BT group included five males and five females with a mean age of 28.5 years (range 24-37 years).

Clinical Measurements

Comparing different groups for clinical measurements presented in **Table. 1** showed that for buccal and palatal bone height loss, there was a significant difference between different groups with the alloplast group having significantly higher bone loss values than Auto-BT (p<0.05), while no statistically significant difference was observed between allograft and the other two groups. For

TABLE (1) Clinical analysis (Buccal and Palatal bone height loss and Bone width loss)

Measurement -	(Mean±SD) (mm)			f-value	p-value
	Allograft	Alloplast	Autogenous tooth bone graft	1-value	p-value
Buccal bone height loss	1.40±0.97 ^{AB}	1.98±0.55 ^A	0.80±0.67 ^B	6.22	0.006*
Palatal bone height loss	1.20±0.88 ^{AB}	2.07±0.83 ^A	0.77 ± 0.46^{B}	3.65	0.039*
Bone width loss	1.42±0.94 ^A	1.95±0.10 ^A	1.05±0.83 ^A	1.43	0.256

the loss in bone width, the difference was not statistically significant (p=0.256).

Radiographic Measurements

Radiographic measurements presented in **Table. 2** showed that for bone width loss there was a significant difference between groups, with β -TCP having significantly higher loss value than other groups (p=0.001). For height loss, the difference was also significant, with the β -TCP group having a significantly higher value than the Auto-BT group (p=0.009). For bone density gain, there was a signifi-

icant difference with the β -TCP group having a significantly lower value than other groups (p<0.001).

Histological Results and Histomorphometric Analysis

Histologically, all groups showed variable amounts of woven bone and lamellar bone with haversian system. Residual graft was detected in which some of the graft remnants were found fused to the newly formed bone especially in the allograft and Auto-BT groups. Osteoblastic rimming was observed lining the graft material and the newly formed bone in the allograft group (Fig.3).

TABLE. (2) Radiographic analysis (Width loss, Height loss and Density gain between different study groups)

Measurement -	(Mean±SD)			C 1	1
	Allograft	Alloplast	Autogenous tooth bone graft	f-value	p-value
Width loss (mm)	1.33±0.05 ^B	1.59±0.16 ^A	1.22±0.31 ^B	8.47	0.001*
Height loss (mm)	1.64 ± 0.38^{AB}	2.00±0.94 ^A	0.88 ± 0.84^{B}	5.68	0.009*
Density gain (HU)	77.04±13.18 ^A	52.69±0.75 ^B	85.99±9.62 ^A	33.38	<0.001*

Means with different superscript letters within the same horizontal row are significantly different *significant (p<0.05)

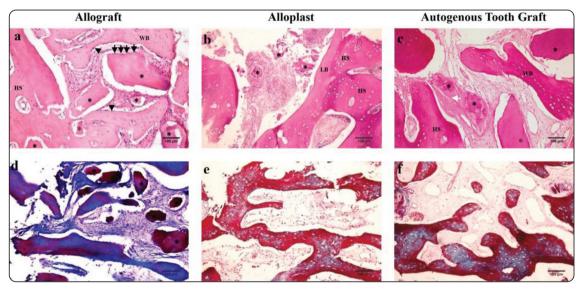


Fig. (3) Photomicrographs of (a,b,c) H& E sections, and (d,e,f) MT sections of biopsy samples (magnification, 200X) showing: Residual graft material in all groups (astrict), (a) Erosion of the surface of allograft residual graft with osteoblastic rimming (black arrows), osteoblastic rimming of the newly formed woven bone trabeculae (black arrow heads), and Haversian system (b) Lamellar bone with Haversian system and woven bone in alloplast group (c) Woven bone and lamellar bone with Haverian system in autogenous tooth graft group, (a and c) fusion of the newly formed bone with the graft material (white arrows). Bone specimens of experimental groups stained with MT showing immature collagen detected by blue color and mature collagen detected with red color, (d) Allograft group showed more immature blue stained collagen than mature collagen, (e,f) Alloplast and autogenous tooth graft groups showed more red mature than immature blue collagen. WB: Woven bone, LB: Lamellar bone, HS: Haversian system

(310) E.D.J. Vol. 71, No. 1 Hadeel Gamal, et al.

The measurements presented in **Table.3** revealed a significant difference in mature bone area percentage between different groups, with the Auto-BT group having the highest value followed by β -TCP and with allograft group having the lowest value (p<0.001). The percentage of mature to immature bone was found to be statistically significant in alloplast and Auto-BT groups.

Moreover, regarding the mean area percentage of the residual graft material, our results showed that there was a significant difference between the tested groups (p<0.001). The highest value was found in the allograft group, followed by the Auto-BT group, while the lowest value was found in the alloplast group. All pairwise comparisons were statistically significant (p<0.001) **Table.4**.

TABLE. (3) Histological analysis (Mean Area % of Mature and immature collagen of bone within and between different study groups)

Measurement -	(Mean±SD) (%)			C 1	1
	Allograft	Alloplast	Autogenous tooth bone graft	f-value	p-value
Mature collagen	11.54±1.14 ^c	21.84±1.81 ^B	24.26±1.95 ^A	162.93	<0.001*
Immature collagen	13.87±3.67 ^A	8.01±0.26 ^C	10.39±1.89 ^B	15.19	<0.001*
t-value	2.00	25.69	25.08		
p-value	0.076	<0.001*	<0.001*		

Means with different superscript letters within the same horizontal row are significantly different *significant (p<0.05)

TABLE (4) Histological analysis (Mean Area % of residual graft material)

Res	idual graft material (Me	£1	p-value	
Allograft	Alloplast	Autogenous tooth bone graft	f-value s tooth bone graft	
8.20±2.48 ^A	1.79±0.35 ^c	4.25±0.75 ^B	69.61	<0.001*

Means with different superscript letters within the same horizontal row are significantly different *significant (p<0.05)

DISCUSSION

The reduction in alveolar ridge dimensions and mucosal thickness after tooth extraction is an inevitable event that could be compensated by ARP [4, 38, 39].

The superiority of flapless extraction of teeth lies in the preservation of the blood supply to the buccal bone, as it does not cause any periosteal detachment. This type of extraction proved to successfully conserve the hard tissue dimensions and keep more healthy keratinized gingival tissue [9,40].

Collagen membrane was chosen in this study, as it can keep soft tissues away from filling the extraction bony defect, allowing the specialized cells with osteogenic ability to regenerate the defect lost tissues. Collagen membranes are resorbable membranes having the advantage of self-degradation with no need for a second surgery to remove [41-43].

The effectiveness of collagen membranes and bone replacements in ARP has been assessed in a number of trials. One study, for example, found that the absorbable collagen membrane and deproteinized bovine bone graft helped preserve the alveolar ridge bone while having no negative effects on the osseointegration of delayed implants [44]. Another trial aimed to reduce the dimensional changes in the alveolar bone post-tooth extraction by using an equine collagen membrane and a collagen cone, suggesting the potential of collagen materials in ARP [45].

The 3 months healing period was selected as it was previously demonstrated that the ideal healing time for a tooth socket is 12 weeks ^[5]. Moreover, after 3 months, the bone formation is adequate for implant placement, as the majority of the graft material is usually substituted by mature bone at that time point ^[9,46]. Additionally, Jeong et al. ^[47] and Kim et al. ^[48] observed the resorption of the graft in their studies after 3 to 6 months of grafting.

Radiographically, CBCT was chosen for the analysis of changes in osseous dimensions and density. With low radiation doses, CBCT produces a more economical and efficient images [49]. MT stain was used for the histomorphometric analysis as it can differentiate between mature and immature collagen in the newly formed bone [50].

Results of the clinical measurements showed that the alloplast group had the highest value of bone height loss than the Auto-BT group, while the Auto-BT group showed the least loss in bone width. In their study Joshi et al., compared ARP using Auto-BT versus β-TCP. After 4 months posttreatment, the vertical and horizontal bone loss was the highest in the ungrafted sites followed by β-TCP sites, and was the least for Auto-BT -grafted sites with a statistically significant difference between the groups [51]. In accordance with our study, Jambhekar et al. [9] reported that allografts resulted in less affection of socket dimensions compared to alloplasts using a cut off healing period of 3 months. This was explained by the ability of the FDBA to regenerate bone or induce new bone formation^[9]. The process of freeze –drying lowers the antigenicity^[52]

and the presence of bone morphogenic proteins exerts an osteoinductive property [5].

Regarding CBCT results, Auto-BT group showed less mean percent decrease in height and width and more density gain with a statistically significant difference between the groups. Similarly, Joshi et al. ^[51] reported that Auto-BT-grafted sites showed less reduction in ridge height and width, which was significantly lower when compared to β-TCP-grafted sites. In a previous study, Jun et al. ^[53] claimed no statistically significant difference in bone density gain and bone height change between Auto-BT and bovine bone graft. Thus, they recommended Auto-BT as a good alternative to other bone graft materials in sinus bone grafting.

Histological evaluation and histomorphometric analysis of the core biopsies obtained 3 months after socket grafting showed variable amounts of new bone in the grafted site of all groups. The presence of osteoblastic layer at the newly formed bone surface and at the surface of the resorbing graft particles of the allograft group indicates the presence of active continuous mineralization. The maturation of bone in all groups was indicated by the uniform osteocytic distribution and Haversian canals [54]. Our results revealed the highest value of mature bone in the Auto-BT group followed by alloplast, while the allograft had the lowest value. This agreed with Menetti et al.'s previous histological study where some of the dental granules were resorbed while others were still present. The newly formed bone was closely connected with the Auto-BT material and the granules were totally surrounded in some areas by the new bone [22]. Additionally, our results go along with that of Joshi et al. [51] who detected less bone and less angiogenesis besides more inflammation in β-TCP-grafted sites when compared to Auto-BT. Moreover, a systematic review on socket grafting by allografts, alloplasts and xenografts stated that alloplasts showed the (312) E.D.J. Vol. 71, No. 1 Hadeel Gamal, et al.

highest percentage of vital bone formation and the lowest percentage of residual graft material (45.53%, 13.67%, respectively) while allografts presented the lowest percentage of vital bone and the highest remnant graft (29.93%,21.75%, respectively). This was explained by the osteoconductive potential and the rapid rate of resorption of the graft material owing to the microporosity in β -TCP ^[9,55].

The properties of Auto-BT explain the best results of this graft material. First, Auto-BT is similar to bone composition in the presence of 70% hydroxiapatite and other calcium phosphate minerals [56]. Additionally, the bioactive bone morphogenic protein 2 and fibroblast growth factor in tooth structure contribute to the osteoinductive and osteoconductive property of Auto-BT [57]. Moreover, the mechanical stability of dentine particles and their ability to firmly integrate with the new bone creating a rigid anchorage site for implant, give the Auto-BT supremacy over other graft materials [58,59].

CONCLUSION

Taken together, we conclude that grafts derived from an extracted human tooth could be considered a good alternative to allografts or alloplasts in alveolar ridge preservation as confirmed clinically, radiographically, and histologically. Autogenous tooth graft represents a good model of recycling autogenous tissues instead of discarding them as medical waste.

Funding

This research did not receive any specific grant for funding agents in the public commercial or notfor-profit sectors.

Conflict of Interest

The authors declare that they have no competing interest

Abbreviations

ARP, Alveolar ridge preservation; RCT, Randomized controlled trials; GBR, Guided bone regeneration; FDBA, freeze-dried bone allografts; β-TCP, beta tricalcium phosphate;

Auto-BT, autogenous tooth bone graft; PDL, Periodontal ligament; PBS, Phosphate-buffered saline; EDTA, Ethylene diamine tetra-acetic acid; H&E, Hematoxylin and Eosin; MT, Masson trichrome special stain; SDG, Smart Dentin Grinder; CBCT, Cone beam computed tomography

REFERENCES

- Schropp L, Wenzel A, Kostopoulos L, Karring T. Bone healing and soft tissue contour changes following singletooth extraction: a clinical and radiographic 12-month prospective study. Int J Period Restor Dent 2003; 23: 313-23. DOI: https://pubmed.ncbi.nlm.nih.gov/12956475/
- Tan WL, Wong TL., Wong MC., Lang NP. A systematic review of post-extractional alveolar hard and soft tissue dimensional changes in humans. Clin Oral Implants Res 2012; 5:1-21. DOI: https://doi.org/10.1111/j.1600-0501.2011.02375.x
- Vignoletti F, Matesanz P, Rodrigo D, Figuero E, Martin C, Sanz M. Surgical protocols for ridge preservation after tooth extraction. A systematic review. Clin Oral Implants Res 2012; 23:22–38. DOI: https://doi.org/10.1111/j.1600-0501.2011.02331.x
- Avila-Ortiz G, Elangovan S, Kramer KWO, Blanchette D, Dawson DV. Effect of alveolar ridge preservation after tooth extraction: a systematic review and metaanalysis. J Dent Res 2014; 93: 950-8. DOI: https://doi. org/10.1177/0022034514541127
- Chan HL, Lin GH, Fu JH, Wang HL. Alterations in bone quality after socket preservation with grafting materials: a systematic review. Int. J. Oral Maxillofac. Implants 2013; 28: 710–20. DOI: https://doi.org/10.11607/jomi.2913
- Chappuis V, Engel O, Reyes M, Shahim K., Nolte LP, Buser D. Ridge alterations post-extraction in the esthetic zone:
 a 3D analysis with CBCT. J. Dent. Res 2013;92: 195S-201S. DOI: https://doi.org/10.1177/0022034513506713

- Atieh MA, Alsabeeha NH, Payne AG, Duncan W, Faggion CM, Esposito M. Interventions for replacing missing teeth: alveolar ridge preservation techniques for dental implant site development. *Cochrane Database of Systematic Re*views, (4). 2021. DOI: https://doi.org/10.1002/14651858. cd010176.pub2
- Kalsi AS, Kalsi JS, Bassi S. Alveolar ridge preservation: why, when, and how. Br Dent J 2019; 227: 264-74. DOI: https://doi.org/10.1038/s41415-019-0647-2
- Jambhekar S, Florian K, Avinash SB. Clinical and histologic outcomes of socket grafting after flapless tooth extraction: a systematic review of randomized controlled clinical trials." J Prosthet Dent 2015; 113: 371-382. DOI: https://doi.org/10.1016/j.prosdent.2014.12.009
- AlGhamdi AS, Shibly O, Ciancio SG. Osseous grafting part I: Autografts and allografts for periodontal regeneration—A literature review. J Int Acad Periodontol 2010; 12: 34-38. DOI: https://pubmed.ncbi.nlm.nih.gov/20465029/
- Eskow AJ, Mealey BL. Evaluation of healing following tooth extraction with ridge preservation using cortical vs. cancellous freeze dried bone allograft. J Periodontol 2014; 85:514–524. DOI: https://doi.org/10.1902/ jop.2013.130178
- 12. Harel N, Moses O, Palti A, Ormianer Z. Long-term results of implants immediately placed into extraction sockets grafted with β -tricalcium phosphate: a retrospective study. J. Oral Maxillofac. Surg 2013; 71: e63–e68. DOI: https://doi.org/10.1016/j.joms.2012.09.022
- 13. Kakar A., Rao B H., Hegde S, et al. Ridge preservation using an in situ hardening biphasic calcium phosphate (β-TCP/HA) bone graft substitute—a clinical, radiological, and histological study. Int. J. Implant Dent 2017; 3: 25. DOI: https://doi.org/10.1186/s40729-017-0086-2
- Darby I. Periodontal materials. Aust. Dent. J 2011;
 107-18. DOI: https://doi.org/10.1111/j.1834-7819.2010.01301.x
- Kim YK, Lee JH, Um IW, Cho WJ. Guided Bone Regeneration Using Demineralized Dentin Matrix: Long-Term Follow-Up. J Oral Maxillofac Surg 2016;74:515.e1-9. DOI: https://doi.org/10.1016/j.joms.2015.10.030
- Cenicante J, Botelho J, Machado V, Mendes JJ, Mascarenhas P, AlcoforadoG, Santos A. The use of autogenous teeth for alveolar ridge preservation:a literature review.
 Appl Sci. 2021;11(4):1853. DOI: https://doi.org/10.3390/app11041853

- Minetti E, Taschieri S, Corbella S. Autologous deciduous tooth-derived material for alveolar ridge preservation: a clinical and histological case report. Case Rep Dent. 2020;2020: 2936878. DOI: https://doi.org/10.1155/2020/2936878
- 18. Santos A, Botelho J, Machado V, Borrecho G, Proenca L, Mendes JJ, Mascarenhas P, Alcoforado G. Autogenous mineralized dentin versus xenograft granules in ridge preservation for delayed implantation inpost-extraction sites: a randomized controlled clinical trial with an 18months follow-up. Clin Oral Implants Res 2021; 32(8):905–915. DOI: https://doi.org/10.1111/clr.13765
- Park M, Mah YJ, Kim DH, Kim ES, Park EJ. Demineralized deciduous tooth as a source of bone graft material: its biological and physicochemical characteristics. Oral Surg Oral Med Oral Pathol Oral Radiol 2015; 120: 307-314. DOI: https://doi.org/10.1016/j.oooo.2015.05.021
- 20. Koga T, Minamizato T, Kawai Y, Miura K., I T, Nakatani Y, et al. Bone Regeneration Using Dentin Matrix Depends on the Degree of Demineralization and Particle Size. PLoS One 2016;11: e0147235. DOI: https://journals.plos.org/plosone/article?id=10.1371/journal.pone.0147235
- 21. Min BM. Oral Biochemistry. DaehanNarae Pub Co. Seoul; 2007.pp. 22-26.
- Minetti E, Corbella S, Taschieri S, Caullo L. Tooth as graft material: Histologic study. Clin. Implant Dent. Relat. Res 2022; 24: 488-496. DOI: https://onlinelibrary.wiley.com/ doi/abs/10.1111/cid.13097
- Kim TW, Seo EW, Song SI. Open reduction and internal fixation of mandibular fracture in an 11-month-old infant: a case report. J Korean Assoc Oral Maxillofac Surg 2013; 39(2):90–93. DOI: https://doi.org/10.5125/jkaoms.2013.39.2.90
- 24. Kim Y., Lee JK, Kim KW, Um IW, Murata M. Healing Mechanism and Clinical Application of Autogenous Tooth Bone Graft Material. Advances in Biomaterials Science and Biomedical Applications. 2013; 405-36. DOI: http:// dx.doi.org/10.5772/53200
- Atiya BK, Shanmuhasuntharam P, Huat S, Abdulrazzak S, Oon H. Liquid nitrogen-treated autogenous dentin as bone substitute: an experimental study in a rabbit model. Int J Oral Maxillofac Implants 2014; 29: 165–170. DOI: https://doi.org/10.11607/jomi.te54
- 26. Wood RA, Mealey BL. Histologic comparison of healing after tooth extractionwith ridge preservation using

(314) E.D.J. Vol. 71, No. 1 Hadeel Gamal, et al.

- mineralized versus demineralizedfreeze-dried bone allograft. J Periodontol 2012;83(3):329–336. DOI: https://doi.org/10.1902/jop.2011.110270
- Wennerberg A, Albrektsson T, Lindhe J. Surface topography of titanium dental implants. En: Lindhe J, Karring T, Lang N, eds. Clinical Periodontology and Implant Dentistry. Blackwell Munksgaard, 2003: 821-828.
- Kim YK, Pang KM, Yun PY, Leem DH, Um IW. Longterm follow-up ofautogenous tooth bone graft blocks with dental implants. Clin Case Rep 2017;5(2):108–118. DOI: https://doi.org/10.1002/ccr3.754
- Shayar A. Autogenous Tooth Graft: An Overview and Meta-Analysis. J Oral Med and Dent Res 2023; 4(2):1 DOI: http://dx.doi.org/10.52793/JOMDR.2023.4(2)-42
- Marei H Fi, Ahmed MG. "Socket preservation for dental implant site development A randomized controlled clinical trial." Egypt. Dent. J 2017; 63 (3-July (Oral Surgery)): 2281-2288. DOI: https://edj.journals.ekb.eg/article 75765.html
- Faul F, Erdfelder E., Lang AG, Buchner A. "G* Power 3: A flexible statistical power analysis program for the social, behavioral, and biomedical sciences. Behav Res Method 2007; 39: 175-191. DOI: https://doi.org/10.3758/BF03193146
- 32. Glick M, Greenberg MS, Ship JA. Introduction to oral medicine and oral diagnosis: evaluation of the dental patient. In: Burket's Oral Medicine. 11th ed. Bc Decker Inc, Hamilton; 2008 pp. 1-6. DOI: https://books-library.net/ files/download-pdf-ebooks.org-1519307052Nl3O9.pdf
- Madhan S, Govindaswamy G, Madhumati S, Singh T. Comparison of Ability of Platelet-rich Fibrin vs CollaPlug in maintaining the Buccal Bone Height of Sockets following Extractions in 20 Patients. JHSR 2017; 8: 1-6. DOI: http://www.johsr.com/doi/JOHSR/pdf/10.5005/jp-journals-10042
- Hauser F, Gaydarov N, Badoud I, Vazquez L, Bernard JP, Ammann P. Clinical and histological evaluation of post extraction platelet-rich fibrin socket filling: a prospective randomized controlled study. Implant Dent 2013; 22: 295– 303. DOI: https://doi.org/10.1097/id.0b013e3182906eb3
- Binderman I, Hallel G, Nardy C, Yaffe A, Sapoznikov L.Processing extracted teeth for immediate grafting of autogenous dentin. Implant Pract US 2015; 2: 43-46.
 DOI: https://www.implantpracticeus.com/wp-content/up-loads/2015/05/43-46_CE-Binderman.pdf

- Rhodes A. Fixation of tissues in: Bancroft JD., Gamble M., Sauvarna S (Eds.) Theory and practice of histological techniques 6th ed. Churchill Livingstone publishers, Philadelphia; 2008. pp. 69-94.
- 37. Bancroft JD, Layton C. Connective and mesenchymal tissues with their stains, in: Bancroft, J.D., Gamble, M., Sauvarna S (Eds.) Theory and practice of histological techniques 6th ed. Churchill Livingstone publishers, Philadelphia; 2008 .pp.187-214.
- 38. Thoma DS, Benic GI, Zwahlen M, Hammerle CH, Jung RE. A systematic review assessing soft tissue augmentation techniques. Clin Oral Implants Res 2009; 4: 146–165. DOI: https://doi.org/10.1111/j.1600-0501.2009.01784.x
- Farmer M, Darby I. Ridge dimensional changes following single-tooth extraction in the aesthetic zone. Clin Oral Implants Res 2014; 25: 272–277. DOI: https://doi.org/10.1111/clr.12108
- 40. Barone A, Borgia V, Covani U, Ricci M, Piattelli A, Iezzi G. Flap versus flapless procedure for ridge preservation in alveolar extraction sockets: a histological evaluation in a randomized clinical trial. Clin. Oral Implants Res 2015; 26: 806-813. DOI: https://doi.org/10.1111/clr.12358
- Wang HL, Carroll MJ. Guided bone regeneration using bone grafts and collagen membranes. Quintessence Int 2001; 32: 504-515. DOI: https://pubmed.ncbi.nlm.nih. gov/11495562/
- Barone A, Aldini NN, Fini M, Giardino R, Calvo Guiradon JL, Covani U. Xenograft versus extraction alone for ridge preservation after tooth removal: a clinical and histomorphometric study. J Periodontol 2008; 79: 1370-1377. DOI: https://doi.org/10.1902/jop.2008.070628
- 43. Retzepi M, Donos N. Guided Bone Regeneration: biological principle and therapeutic applications. Clin. Oral Implants Res 2010; 21: 567-576. DOI: https://doi.org/10.1111/j.1600-0501.2010.01922.x
- 44. Pang C, Ding Y, Zhou H, Qin R, Hou R, Zhang G, Hu K. Alveolar ridge preservation with deproteinized bovine bone graft and collagen membrane and delayed implants. J Craniofac Surg 2014; 25(5):1698–1702. DOI: https://doi.org/10.1097/scs.00000000000000887
- Schnutenhaus S, Doering I, Dreyhaupt J, Rudolph H, Luthardt RG (2018) Alveolar ridge preservation with a collagen material: a randomizedcontrolled trial. J Periodontal Implant Sci 48(4):236–250. DOI: https://doi.org/10.5051/jpis.2018.48.4.236

- 46. Neiva R, Pangi G, Duarte F, Park CH, Yi E, Holman LA, et al. Analysis of tissue neogenesis in extraction sockets treated with guided bone regeneration: clinical, histological, and micro-CT results. Int J Periodontics Restorative Dent 2011; 31: 457-769. DOI: https://pubmed.ncbi.nlm.nih.gov/21845241/
- 47. Jeong KI, Kim SG, Oh JS, Lim SC. Maxillary sinus augmentation using autogenous teeth: preliminary report. J Korean Assoc Maxillofac Plast Reconstr Surg 2011; 33: 256-263. DOI: https://www.koreamed.org/SearchBasic.php?RID=1449177
- Kim YK, Lee HJ, Kim KW, Kim SG, Um IW. Guided bone regeneration using autogenous teeth: case reports. J. Korean Assoc. Oral Maxillofac. Surg 2011; 37:142-147. DOI: https://synapse.koreamed.org/articles/1032460
- Sukovic P. Cone beam computed tomography in craniofacial imaging. Orthod. Craniofacial. Res 2003; 6: 31-36.
 DOI: https://doi.org/10.1034/j.1600-0544.2003.259.x
- 50. Suvik A, Effendy AWM. The use of modified Masson's trichrome staining in collagen evaluation in wound healing study. Mal J Vet. Res 2012; 3: 39-47. DOI: https://www. dvs.gov.my/dvs/resources/user_15/Mjvr%20v3/MJVR-V3N1-p39-47-email.pdf
- Joshi CP, Dani NH, Khedkar SU. Alveolar ridge preservation using autogenous tooth graft versus beta-tricalcium phosphate alloplast: A randomized, controlled, prospective, clinical pilot study. J Indian Soc Periodontol 2016; 20: 429-434. DOI: https://doi.org/10.4103/0972-124x.188335
- 52. Hanser T, Khoury F. Extraction site management in the esthetic zone using autogenous hard and soft tissue grafts: a 5- year consecutive clinical study. Int J Periodontics Restorative Dent 2014; 34: 305-312. DOI: https://doi.org/10.11607/prd.1749

- Jun SH, Ahn JS, Lee JI, Ahn KJ, Yun PY, Kim YK. A prospective study on the effectiveness of newly developed autogenous tooth bone graft material for sinus bone graft procedure. J Adv Prosthodont 2014; 6: 528-538. DOI: https://doi.org/10.4047/jap.2014.6.6.528
- 54. Adel-Khattab D, Afifi NS, El Sadat SM, Aboul-Fotouh MN, Tarek K, Horowitz RA. Bone regeneration and graft material resorption in extraction sockets grafted with bioactive silica-calcium phosphate composite versus non-grafted sockets: clinical, radiographic, and histological findings. JPIS 2020; 50(6):418. DOI: https://doi.org/10.5051/jpis.2000040002
- 55. Horowitz R, Holtzclaw D, Rosen PS. A review on alveolar ridge preservation following tooth extraction. J Evid Based Dent Pract 2012; 12: 149-160. DOI: https://doi.org/10.1016/s1532-3382(12)70029-5
- Kim YK, Kim SG, Oh JS, et al. Analysis of the inorganic component of autogenous tooth bone graft material. J Nanosci Nanotechnol 2011; 11: 7442-7445. DOI: https://doi. org/10.1166/jnn.2011.4857
- Nampo T, Watahiki J, Enomoto A, Taguchi T, Ono M, Nakano H, et al. A new method for alveolar bone repair using extracted teeth for the graft material. J Periodontol 2010; 8: 1264-1272. DOI: https://doi.org/10.1902/ jop.2010.100016
- 58. Andersson L. Dentin xenografts to experimental bone defects in rabbit tibia are ankylosed and undergo osseous replacement. Dent. Traumatol 2010; 26: 398-402. DOI: https://doi.org/10.1111/j.1600-9657.2010.00912.x
- Binderman I, Hallel G, Nardy C, Yaffe A, Sapoznikov L. A novel procedure to process extracted teeth for immediate grafting of autogenous dentin. J Interdiscipl Med Dent Sci 2014; 2: 2. DOI: https://puredent.dk/pdf/sdg/paperdentinsocket.pdf